

ELI.H.A *Schistosoma*

Serodiagnosis of bilharzia by indirect haemagglutination

120 tests
(Ref. 66600)

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1 - AIM

ELI.H.A *Schistosoma* allows the quantitative determination, by indirect haemagglutination, of serum antibodies from patients with bilharziasis caused by *Schistosoma mansoni* (intestinal form) and *Schistosoma haematobium* (urinary form).

Each kit allows 120 tests to be carried out or 20 reactions of 6 dilutions.

2 - INTRODUCTION

The bilharzias, or schistosomiasis, represent a group of parasitic diseases caused by the non-segmented flat worms of the genus *Schistosoma*.

Immunity is acquired progressively in the presence of adult worms. It is however the schistosome eggs that are the pathogenic element of the disease. The disease proceeds in three successive phases: contamination - invasion - disease state.

The diagnosis of bilharzia can be made by identification of the eggs, but these cannot be detected before the "disease state" phase. Parasite immunological diagnosis is thus essential in the initial stages of this disease.

3 - PRINCIPLE

ELI.H.A *Schistosoma* is based on the indirect haemagglutination principle. The sensitized red blood cells consist of sheep red blood cells covered with a *Schistosoma mansoni* antigen.

The presence of specific serum antibodies results in agglutination of the sensitized red blood cells resulting in a cloudy red/brown deposit coating the well. In the absence of specific antibodies, the red blood cells form a ring-like deposit at the bottom of the well.

The non-sensitized red blood cells ensure the specificity of the reaction making it possible to eliminate any interference from the natural anti-sheep agglutinins (Forssman heteroantibodies, infectious mononucleosis antibodies...).

The reaction is carried out in a U-microplate.

Handling is simple and fast, with results within 2 hours.

4 - REAGENTS AND MATERIAL

Description	Quantity
R1: Vial of 2.4 mL of sensitized red blood cells	1
R2: Vial of 1 mL of non-sensitized red blood cells	1
BUF: Vial of 55 mL of phosphate buffer pH 7.2	1
R3: Vial of 2 mL of adsorbent	1
CONTROL +: Vial of 0.2 mL of titrated positive control	1
CONTROL -: Vial of 0.2 mL of negative control	1
MICROPLATE: Microplate with a U-bottom	2
DROPPER: Special dropper	2

5 - PRECAUTIONS

- The reagents are intended for *in vitro* diagnostic use only and must be handled by authorized personnel. Tests are for a single use only.
- All the reagents, except the **BUF** reagent, contain raw materials of animal origin and must be handled with caution.
- Patient samples are potentially infectious. They must be handled with caution, in observance of hygiene rules and the current regulations for this type of product in the country of use.
- The **CONTROL** reagents contain sodium azide (<0.1%).
- Do not use reagents after the expiry date.
- Do not use reagents from different batch numbers.
- Prior to use, allow the serum and the reagents to reach room temperature.
- Carefully shake the **R1** and **R2** reagents before use.
- When dispensing the **R1** and **R2** reagents, make sure that the dropper is perfectly vertical. Check for the absence of air bubbles in the drops to ensure constant delivery volumes.

6 - SAMPLE COLLECTION AND TREATMENT

Use fresh serum or serum preserved at -20°C, and not showing any sign of haemolysis, cloudiness or of contamination.

Avoid repeated freezing and defrosting.

Do not decompartment the serum.

7 - STABILITY, STORAGE AND PREPARATION OF REAGENTS

The reagents are ready-to-use.

All the reagents stored at 2-8°C, in their original packaging, are stable until the expiry date indicated on the box. Do not freeze.

8 - MATERIAL REQUIRED BUT NOT SUPPLIED

- Automatic pipette(s) with a pipetting volume adapted to the volume that will be measured;
- Contaminated waste containers;
- Centrifuge;
- Haemolysis tubes.

9 - METHOD

Allow the reagents to reach room temperature before use.

9.1 - Sample preparation

Carry out a 1:40 dilution of the serum to be tested:

- 50 µL of serum;
- 1.95 mL of **BUF** reagent.

9.2 - Realization of the test on a microplate

- Using a multichannel micropipette, add 50 µL of **BUF** reagent to 8 wells of the microplate.

- Using a micropipette, add 50 µL of diluted serum to the 1st well.

Mix the serum with the **BUF** reagent and carry out a serial dilution, preferably using a microdiluter, by transferring 50 µL from the 1st well into the 2nd, then 50 µL from the 2nd to the 3rd, and so on until the 6th well is reached. 50 µL from the 6th well is then discarded.

In this way, dilutions from 1:80 to 1:2560 are obtained.

- Add 50 µL of diluted serum to the 7th well.

Mix the serum with the **BUF** reagent and then discard 50 µL.

This dilution (1:80) is the serum control, whose role is to detect the natural anti-sheep agglutinins that could be present in certain serum samples.

- Carefully shake the **R1** and **R2** reagents.

- Add 1 drop of **R1** reagent to the first 6 wells.
- Add 1 drop of **R2** reagent to the 7th well (serum control).
- Add 1 drop of **R1** reagent to the 8th well (reagent control) whose role is to control the validity of the **BUF** and **R1** reagent.

Note: Only carry out one reagent control for each series of tests.

- Very carefully, shake the contents of the wells:

- either manually, by tapping laterally the side of the microplate that has been posed flat on the bench;
- or by using a vibrating plate shaker for microtiter plates (for example at 1300 rpm for 10 seconds). Do not use an orbital shaker.

- Now leave the plate to rest, away from any sources of vibration.

- The plate can be read after 2 hours.

9.3 - Adsorption of the natural anti-sheep agglutinins in the event of agglutination of the serum control

- Carefully shake the **R3** reagent.

- In a tube, add and mix:

- 0.1 mL of serum;
- 0.3 mL of **R3** reagent.

- Incubate at room temperature for 60 minutes.

- Centrifuge at 2000 rpm for 15 minutes.

- Collect the supernatant; the serum is now at a 1:4 dilution.

- Carry out a 1:10 dilution of the supernatant in **BUF** reagent to obtain an adsorbed stock dilution (1:40).

- Follow the steps described in "Realization of the test on a microplate", but replace the stock dilution by the adsorbed stock dilution.

10 - READING

Negative reaction:

Absence of haemagglutination.

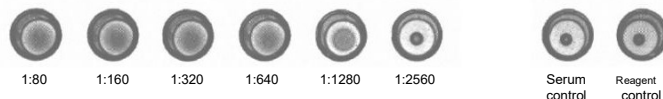
Presence of a more or less large ring at the bottom of the well.

Positive reaction:

Presence of haemagglutination.

Presence of a cloudy red/brown deposit coating the well, sometimes there is the presence of a fine peripheral border.

Example: Serum positive at a dilution of 1:1280



11 - INTERPRETATION OF RESULTS

Titer < 1:160:

Non significant reaction of a progressive infection.

Can correspond to a previous or treated infection.
Renew the test 2 to 3 weeks later and also carry out an electrosynthesis or an immunoelectrophoresis test.

Titer > 1:160:

Significant reaction.

Presumption of an active infection.

12 - INTERNAL QUALITY CONTROL

The **CONTROL +** and **CONTROL -** reagents must be treated like test serums. The titer of the **CONTROL +** reagent must be the same as the titer printed on the vial label \pm one dilution. There must not be any haemagglutination of the **CONTROL -**. If haemagglutination is present then the test is not valid.

13 - CAUSES OF ERROR AND TEST LIMITS

- Poor conservation of the serum.
- Poor conservation of the reagents after opening.
- Only use the droppers provided in the kit.
- Do not interchange the droppers between the **R1** and **R2** reagents.
- In the case of a positive reaction in the first 6 wells, carry out a further serial dilution in order to determine the titer limit of haemagglutination.
- The serum control must give a negative reaction (ring). In the event of haemagglutination of this control, it will be necessary to renew the test after having eliminated the natural anti-sheep agglutinins from the serum by adsorption.
- The reagent control must give a negative reaction (ring). In the event of haemagglutination of this control, the **ELI.H.A Schistosoma** cannot be used.
- Certain serums, whose antibody concentration is very high, can give rise to a zone phenomenon (with disappearance of the clouding) in the initial dilutions, which disappears in the subsequent dilutions.
- The quality of the reagents makes it possible to carry out the reaction in the evening and to read the test the following morning, provided that the microplate is not moved in any way and is protected from any sources of vibration
- In all cases, it is necessary that the clinical, epidemiologic and biological data are taken fully into consideration before establishing the final diagnosis.

14 - PERFORMANCE

ELI.H.A Schistosoma consists of red blood cells sensitized by a highly purified *Schistosoma mansoni* antigen, which ensures that this indirect haemagglutination test is both sensitive and specific.

The performance evaluations of 63 positive sera (*Schistosoma haematobium* and *mansoni*) and 56 negative sera showed a sensitivity of 85.7% and a specificity of 98.2%.

15 - WASTE ELIMINATION

Waste should be disposed of in accordance with the hygiene rules and current regulations for this kind of product in the country of use.

If the **BUF** reagent is spilled, clean the work area with absorbent paper and rinse with water.

If a serum or another reagent is spilled on the work area, clean using bleach and absorbent paper.

16 - BIBLIOGRAPHY

1. S. HOSHINO, M.-E. CAMARGO, L.-C. DA SILVA - Standardization of a haemagglutination test for schistosomiasis with formalin-treated human erythrocytes - *The American Journal of Tropical Medicine and Hygiene*, Vol. 19, N°3, 463/470.
2. A. CAPRON, J. BIGUET, P. TRAN VAN KY, Y. MOSCHETTO - Immunological studies in various types of schistosomiasis - *Department of Parasitology, Faculty of Medicine, University of Lille*, Lille, France.
3. J. TRIBOULEY, J. TRIBOULEY-DURET, M. APPRIOU, D. BERNARD, R. PAUTRIZEL - Application de la réaction d'hémagglutination passive au diagnostic sérologique de la schistosomiasis à *Schistosoma mansoni* - *Bull. Organ. Mond. Santé*, 1976, Vol. 54, 695/702.
4. L. AHMED, A. DODIN - Valeur de la réaction d'hémagglutination passive dans la bilharziose - *Bulletin de la société de Pathologie Exotique*, 1977, N°5, 485/489.
5. P. AMBROISE-THOMAS, R. GRILLOT - L'hémagglutination indirecte dans le diagnostic des bilharzioses. Comparaison à l'immuno-fluorescence indirecte dans l'étude de 3624 sérums humains - *Bulletin de la société de Pathologie Exotique*, 1980, N°3, 277/288.
6. P. WATTRE, M. CAPRON, J.-P. DESSAINT, A. CAPRON - Apport de l'immunologie au diagnostic et au traitement des maladies parasitaires - *Immunologie (I)* - R.P., 1980, 30, 15.
7. M. EL SAYED AZAB, E. ABBAS EL ZAYAT - Evaluation of purified antigens in haemagglutination test (IHA) for determination of cross reactivities in diagnosis of fascioliasis and schistosomiasis - *Journal of the Egyptian Society of Parasitology*, 1996, Vol 26, N°3, 677-685.